

Procedure & Checklist - Greater Than 10 kb Template Preparation Using AMPure® PB Beads

Before You Begin

This procedure can be used to prepare greater than 10 kb libraries from 5 µg of sheared and concentrated DNA. If preparing larger amounts of DNA, scale all reaction volumes proportionally (e.g., if the input amount of DNA is double the amount set forth in this procedure, then double all the reaction volumes listed in the tables).

If a BluePippin™ size selection system is not available or if the SMRTbell® libraries are less than 500 ng, an alternative method using AMPure PB beads may be used to remove fragments in the 1 kb to 2 kb range. A more aggressive AMPure PB purification, using 0.40X AMPure PB beads (in the final step of the purification process) may also be used. Note that AMPure PB beads do not completely remove all short SMRTbell templates. As a result, the resulting average subread length will be shorter compared to a library size selected with the BluePippin System.

| Insert Size Target | Insert Size Range | Sheared and Concentrated DNA Amount | Ligation |
|--------------------|-------------------|-------------------------------------|----------|
| 10 kb to 20 kb | > 10 kb | 5 µg | Blunt |

Evaluate Genomic DNA Quality

We recommend using a Pulsed Field Electrophoresis system for evaluating gDNA quality. There are two commercially available systems capable of resolving HMW DNA fragments and smears.

- Bio-Rad® CHEF Mapper® XA Pulsed Field Electrophoresis system. The procedure is available [here](#).
- Sage Science's Pippin Pulse Electrophoresis Power Supply. The procedure is available [here](#).

Alternatively, Advanced Analytical Technologies, Inc. FEMTO Pulse™ is an automated pulsed-field capillary electrophoresis instrument for evaluating the integrity of genomic DNA with a run time of approximately 1.5 hours.

Fragment and Concentrate DNA

Use a Covaris® g-TUBE® device to shear your DNA sample. General recommendations for use of g-TUBEs can be found in the g-TUBE user manual available for download from the Covaris website (see <http://covarisinc.com/products/g-tube/>). Note that although Covaris provides recommendations for shearing gDNA to 20 kb, these parameters result in sheared gDNA with average insert sizes of less than 20 kb, and are **NOT** suitable for use in this protocol. To generate sheared gDNA with average insert sizes >20 kb, you **MUST** follow the specific instructions provided here:

1. Dilute DNA to 200-300 ng/µL in Elution Buffer (EB). The sample volume may range from 20-100 µL.
2. Shear at 5500 rpm (2029 x g) for 2 minutes in an Eppendorf® MiniSpin plus.
3. Check for any residual sample remaining in the upper chamber. If present, re-spin for another 2 minutes. Continue spinning until all the sample has passed through the orifice.
4. Invert and spin at 5500 rpm (2029 x g) until all the sample has passed through the orifice.
5. Recover your sample into a 1.5 mL LoBind microcentrifuge tube.

Depending upon the quality of your sample, approximately 20% sample loss is to be expected as a result of the shearing and concentration process. Therefore, be sure to have sufficient amounts of starting DNA in order to have at least 5 µg of sheared and concentrated DNA (140 ng/µL) for the subsequent repair steps.






| STEP | ✓ | Concentrate DNA | Notes |
|------|---|---|-------|
| 1 | | <p>Add 0.45X volume of AMPure PB magnetic beads. _____ μL of sample \times 0.45X = _____ μL of beads</p> <p>Note that the beads must be brought to room temperature before use and all AMPure PB bead purification steps should be performed at room temperature.</p> <p>Before using, mix the bead reagent well until the solution appears homogenous. Pipette the reagent slowly since the bead mixture is viscous and precise volumes are critical to the purification process.</p> | |
| 2 | | Mix bead/DNA solution thoroughly by tapping the tube gently. Do not pipet to mix. | |
| 3 | | Quickly spin down the tube (for 1 second) to collect the beads. | |
| 4 | | Allow the DNA to bind to beads by gentle end-over-end rotation for 15-20 minutes at room temperature. We recommend using a tube rotator. | |
| 5 | | Spin down the tube (for 1 second) to collect beads. | |
| 6 | | Place the tube in a magnetic bead rack until the beads collect to the side of the tube and the solution appears clear. | |
| 7 | | <p>With the tube still on the magnetic bead rack, slowly pipette off cleared supernatant and save in another tube. Avoid disturbing the bead pellet.</p> <p>If the DNA is not recovered at the end of this procedure, you can add equal volumes of AMPure PB beads to the saved supernatant and repeat the AMPure PB bead purification steps to recover the DNA.</p> | |
| 8 | | <p>Wash beads with freshly prepared 70% ethanol.</p> <p>Note that 70% ethanol is hygroscopic and should be prepared FRESH to achieve optimal results. Also, 70% ethanol should be stored in a tightly capped polypropylene tube for no more than 3 days.</p> <ul style="list-style-type: none"> – Do not remove the tube from the magnetic rack. – Use a sufficient volume of 70% ethanol to fill the tube (1.5 mL for 1.5 mL tube or 2 mL for 2 mL tube). Slowly dispense the 70% ethanol against the side of the tube opposite the beads. Let the tube sit for 30 seconds. – Do not disturb the bead pellet. – After 30 seconds, pipette and discard the 70% ethanol. | |
| 9 | | Repeat step 8 above. | |
| 10 | | <p>Remove residual 70% ethanol.</p> <ul style="list-style-type: none"> – Remove tube from magnetic bead rack and spin to pellet beads. Both the beads and any residual 70% ethanol will be at the bottom of the tube. – Place the tube back on magnetic bead rack. – Let beads separate fully. – Pipette off any remaining 70% ethanol. | |

| STEP | ✓ | Concentrate DNA | Notes |
|------|---|---|-------|
| 11 | | Check for any remaining droplets in the tube. If droplets are present, repeat step 10. | |
| 12 | | Remove the tube from the magnetic bead rack and allow beads to air-dry (with the tube caps open) for 30 to 60 seconds. | |
| 13 | | <p>For 5 µg of input sheared gDNA, elute in 37 µL Elution buffer. If you started with more than 5 µg input sheared gDNA, scale volume of EB proportionally (i.e., for 6-10 µg of DNA, elute in 72 µL EB).</p> <p>Add the Pacific Biosciences[®] Elution Buffer volume to your beads. Tap the tube with finger gently to mix, until beads are uniformly resuspended. <u>Do not pipet to mix.</u></p> <ul style="list-style-type: none"> – Elute the DNA by letting the mix stand at room temperature for 2 minutes. – Spin the tube down to pellet beads, then place the tube back on the magnetic bead rack. – Let beads separate fully. Then without disturbing the bead pellet, transfer supernatant to a new 1.5 mL Lo-Bind tube. – Discard the beads. | |
| 14 | | <p>Verify your DNA amount and concentration using a Qubit quantitation platform.</p> <ul style="list-style-type: none"> – Measure the DNA concentration using a Qubit fluorometer. – Using 1 µL of the eluted sample, make a 1:10 dilution in EB. – Use 1 µL of this 1:10 dilution to measure the DNA concentration using a Qubit dsDNA HS Assay kit according to the manufacturer's recommendations. <p>Yield up to this point should be 80%. The remaining 9 µL of 1:10 diluted sample may be used for QC by pulsed-field gel electrophoresis or pulsed-field capillary electrophoresis.</p> | |
| 15 | | The sheared DNA can be stored for up to 24 hours at 4°C or at -20°C for longer duration. | |
| 16 | | Actual recovery per µL and total available sample material: _____ | |

ExoVII Pre-Treatment of DNA

Use the following table to remove single-stranded ends from 5 µg of sheared gDNA. If starting with more than 5 µg of sheared gDNA, scale the reaction volumes proportionally (i.e., for a mass between 6 - 10 µg of DNA scale the total volume to 96 µL).


1. In a LoBind microcentrifuge tube, add the following reagents:

| Reagent | Tube Cap Color | Stock Conc. | Volume | Final Conc. | ✓ | Notes |
|--------------------------|---|-------------|---------|-------------|---|-------|
| Sheared DNA (5 µg) | | | 36.0 µL | | | |
| DNA Damage Repair Buffer |  | 10 X | 5.0 µL | 1 X | | |
| NAD ⁺ |  | 100 X | 0.5 µL | 1 X | | |
| ATP high |  | 10 mM | 5.0 µL | 1 mM | | |
| dNTP |  | 10 mM | 0.5 µL | 0.1 mM | | |
| ExoVII |  | 10 U/µL | 1.0 µL | 0.2 U/µL | | |
| Total Volume | | | 48.0 µL | | | |

2. Mix the reaction well by gently tapping the tube.
3. Spin down contents of tube with a quick spin in a microfuge.
4. Incubate at 37°C for 15 minutes, then return the reaction to 4°C.

Repair DNA Damage


Use the following table to prepare your reaction. For more than 5 µg input DNA, scale all reactions volumes proportionally.

| Reagent | Tube Cap Color | Stock Conc. | Volume | Final Conc. | ✓ | Notes |
|-----------------------|---|-------------|---------|-------------|---|-------|
| DNA (ExoVII treated) | | | 48.0 µL | | | |
| DNA Damage Repair Mix |  | 25 X | 2.0 µL | 1X | | |
| Total Volume | | | 50.0 µL | | | |

1. Mix the reaction well by gently tapping the tube.
2. Spin down contents of tube with a quick spin in a microfuge.
3. Incubate at 37°C for 20-60 minutes, return the reaction to 4°C for 1 to 5 minutes.

Repair Ends

Use the following table to prepare your reaction then purify the DNA. For more than 5 µg input DNA, scale all reactions volumes proportionally.

| Reagent | Tube Cap Color | Stock Conc. | Volume | Final Conc. | ✓ | Notes |
|-----------------------|---|-------------|---------|-------------|---|-------|
| DNA (Damage Repaired) | | | 50.0 µL | | | |
| End Repair Mix |  | 20 X | 2.5 µL | 1X | | |
| Total Volume | | | 52.5 µL | | | |





1. Mix the reaction well by gently tapping the tube.
2. Spin down contents of tube with a quick spin in a microfuge.
3. Incubate at 25°C for 5-10 minutes, return the reaction to 4°C. Proceed to the next step.

| STEP | ✓ | Purify DNA | Notes |
|------|---|--|-------|
| 1 | | Add 0.45X volume of AMPure PB beads to the End-Repair reaction. | |
| 2 | | Mix the bead/DNA solution thoroughly by gently tapping the tube. Do not pipet to mix. | |
| 3 | | Quickly spin down the tube (for 1 second) to collect the beads. Do not pellet beads. | |
| 4 | | Allow the DNA to bind to beads by gentle rotation for 15-20 minutes at room temperature. We recommend using a tube rotator. | |
| 5 | | Spin down the tube (for 1 second) to collect beads. | |
| 6 | | Place the tube in a magnetic bead rack to collect the beads to the side of the tube. | |
| 7 | | Slowly pipette off cleared supernatant and save (in another tube). Avoid disturbing the bead pellet. If the DNA is not recovered at the end of this procedure, you can add equal volumes of AMPure PB beads to the saved supernatant and repeat the AMPure PB bead purification steps to recover the DNA. | |
| 8 | | Wash beads with freshly prepared 70% ethanol. Note that 70% ethanol is hygroscopic and should be prepared FRESH to achieve optimal results. Also, 70% ethanol should be stored in a tightly capped polypropylene tube for no more than 3 days. <ul style="list-style-type: none"> – Do not remove the tube from the magnetic rack. – Use a sufficient volume of 70% ethanol to fill the tube (1.5 mL for 1.5 mL tube or 2 mL for 2 mL tube). Slowly dispense the 70% ethanol against the side of the tube opposite the beads. – Do not disturb the bead pellet. – After 30 seconds, pipette and discard the 70% ethanol. | |
| 9 | | Repeat step 8 above. | |
| 10 | | Remove residual 70% ethanol. <ul style="list-style-type: none"> – Remove tube from magnetic bead rack and spin to pellet beads. Both the beads and any residual 70% ethanol will be at the bottom of the tube. – Place the tube back on magnetic bead rack. – Pipette off any remaining 70% ethanol. | |
| 11 | | Check for any remaining droplets in the tube. If droplets are present, repeat step 10 . | |
| 12 | | Remove the tube from the magnetic bead rack and allow beads to air-dry (with tube caps open) for 30 to 60 seconds. | |
| 13 | | For 5 µg of input sheared gDNA, elute in 23 µL Elution buffer. If you started with more than 5 µg input sheared gDNA, scale volume of EB proportionally (i.e., for 6-10 µg of DNA, elute in 46 µL EB). Add the Pacific Biosciences® Elution Buffer volume to your beads. Tap the tube with finger gently to mix. <u>Do not pipet to mix</u> . <ul style="list-style-type: none"> – Elute the DNA by letting the mix stand at room temperature for 2 minutes. – Spin the tube down to pellet beads, then place the tube back on the magnetic bead rack. – Let beads separate fully. Then without disturbing the bead pellet, transfer supernatant to a new 1.5 ml Lo-Bind tube. – Discard the beads. | |

| | | |
|-----------|---|--|
| 14 | Optional: Verify your DNA amount and concentration using a Qubit quantitation platform. <ul style="list-style-type: none"> – Measure the DNA concentration using a Qubit fluorometer. – Using 1 μL of the eluted sample, make a 1:10 dilution in EB. – Use 1 μL of this 1:10 dilution to measure the DNA concentration using a Qubit fluorometer and the dsDNA HS Assay kit according to the manufacturer's recommendations. – The remaining 9 μL of 1:10 diluted sample may be used for QC by pulsed-field gel electrophoresis or pulse-field capillary electrophoresis. | |
| 15 | The End-Repaired DNA can be stored overnight at 4°C or at -20°C for longer duration. | |
| 16 | Actual recovery per μL and total available sample material: _____ | |

Prepare Blunt Ligation Reaction



Use the following table to prepare your reaction, adding the components below in the order listed. Be sure to mix the DNA and adapter BEFORE adding ligase. For more than 5 μg input DNA, scale all reaction volumes proportionally.

| Reagent | Tube Cap Color | Stock Conc. | Volume | Final Conc. | ✓ | Notes |
|-----------------------------------|---|---------------------|--------------------|-----------------------|---|-------|
| DNA (End Repaired) | | | 23.0 μL | | | |
| Blunt Adapter (20 μM) |  | 20 μM | 10.0 μL | 5 μM | | |
| Mix before proceeding | | | | | | |
| Template Prep Buffer |  | 10 X | 4.0 μL | 1X | | |
| ATP low |  | 1 mM | 2.0 μL | 0.05 mM | | |
| Mix before proceeding | | | | | | |
| Ligase |  | 30 U/ μL | 1.0 μL | 0.75 U/ μL | | |
| Total Volume | | | 40.0 μL | | | |

1. Mix the reaction well by gently tapping the tube.
2. Spin down contents of tube with a quick spin in a microfuge.
3. Incubate at 25°C for 15 – 60 minutes. Ligation reactions can be left at 25 °C overnight.
4. Incubate at 65°C for 10 minutes to inactivate the ligase, then return the reaction to 4°C.

ExoIII/VII Digestion to Remove Failed Ligation Products

Use the following table to prepare your reaction. For more than 5 µg input DNA, scale all reaction volumes proportionally.

| Reagent | Tube Cap Color | Stock Conc. | ✓ | Volume |
|--------------------------------|---|-------------|---|---------|
| Ligated DNA | | | | 40.0 µL |
| Mix reaction well by pipetting | | | | |
| ExoIII |  | 100.0 U/µL | | 1.0 µL |
| ExoVII |  | 10.0 U/µL | | 1.0 µL |
| Total Volume | | | | 42.0 µL |

1. Mix the reaction well by gently tapping the tube.
2. Spin down contents of tube with a quick spin in a microfuge.
3. Incubate at 37°C for 1 hour, then return the reaction to 4°C.
4. You must immediately proceed with AMPure PB bead purification after this step.

Purify SMRTbell Templates

There are 3 purification steps, 2 using 0.45X volume of AMPure PB beads the third using 0.40X volume of AMPure PB beads.

| STEP | ✓ | Purify SMRTbell® Templates - First Purification | Notes |
|------|---|--|-------|
| 1 | | Add 0.45X volume of AMPure PB beads to the exonuclease-treated DNA. | |
| 2 | | Mix bead/DNA solution thoroughly by tapping the tube gently. Do not pipet to mix. | |
| 3 | | Quickly spin down the tube (for 1 second) to collect the beads. | |
| 4 | | Allow the DNA to bind to beads by gentle end-over-end rotation for 15-20 minutes at room temperature. We recommend using a tube rotator. | |
| 5 | | Spin down the tube (for 1 second) to collect beads. | |
| 6 | | Place the tube in a magnetic bead rack until the beads collect to the side of the tube and the solution appears clear. | |
| 7 | | With the tube still on the magnetic bead rack, slowly pipette off cleared supernatant and save in another tube. Avoid disturbing the bead pellet. If the DNA is not recovered at the end of this procedure, you can add equal volumes of AMPure PB beads to the saved supernatant and repeat the AMPure PB bead purification steps to recover the DNA. | |
| 8 | | Wash beads with freshly prepared 70% ethanol. Note that 70% ethanol is hygroscopic and should be prepared FRESH to achieve optimal results. Also, 70% ethanol should be stored in a tightly capped polypropylene tube for no more than 3 days. <ul style="list-style-type: none"> – Do not remove the tube from the magnetic rack. – Use a sufficient volume of 70% ethanol to fill the tube (1.5 mL for 1.5 mL tube or 2 mL for 2 mL tube). Slowly dispense the 70% ethanol against the side of the tube opposite the beads. Let the tube sit for 30 seconds. – Do not disturb the bead pellet. – After 30 seconds, pipette and discard the 70% ethanol. | |
| 9 | | Repeat step 8 above. | |
| 10 | | Remove residual 70% ethanol. <ul style="list-style-type: none"> – Remove tube from magnetic bead rack and spin to pellet beads. Both the beads and any residual 70% ethanol will be at the bottom of the tube. – Place the tube back on magnetic bead rack. – Let beads separate fully. – Pipette off any remaining 70% ethanol. | |
| 11 | | Check for any remaining droplets in the tube. If droplets are present, repeat step 10 . | |
| 12 | | Remove the tube from the magnetic bead rack and allow beads to air-dry (with the tube caps open) for 30 to 60 seconds. | |
| 13 | | For up to 10 µg input SMRTbell library, elute the DNA off the beads in 100 µL of Elution buffer. If you have more than 10 µg library, scale volume of EB proportionally. Add the Pacific Biosciences® Elution Buffer volume to your beads. Tap the tube with finger gently to mix. <u>Do not pipet to mix.</u> <ul style="list-style-type: none"> – Elute the DNA by letting the mix stand at room temperature for 2 minutes. – Spin the tube down to pellet beads, then place the tube back on the magnetic bead rack. – Let beads separate fully. Then without disturbing the bead pellet, transfer supernatant to a new 1.5 ml Lo-Bind tube. – Discard the beads. | |
| 14 | | The eluted DNA in 100 µL Elution Buffer should be taken into the second 0.45X AMPure PB bead purification step. | |

| STEP | ✓ | Purify SMRTbell® Templates - Second Purification | Notes |
|------|---|---|-------|
| 1 | | Add 0.45X volume of AMPure PB beads to the eluted DNA from the first AMPure PB bead purification step above. | |
| 2 | | Mix bead/DNA solution thoroughly by tapping the tube gently. Do not pipet to mix. | |
| 3 | | Quickly spin down the tube (for 1 second) to collect the beads. Do not pellet beads. | |
| 4 | | Allow the DNA to bind to beads by gentle rotation for 15-20 minutes at room temperature. We recommend using a tube rotator. | |
| 5 | | Spin down the tube (for 1 second) to collect beads. | |
| 6 | | Place the tube in a magnetic bead rack to collect the beads to the side of the tube. | |
| 7 | | Slowly pipette off cleared supernatant and save (in another tube). Avoid disturbing the bead pellet. If the DNA is not recovered at the end of this procedure, you can add equal volumes of AMPure PB beads to the saved supernatant and repeat the AMPure PB bead purification steps to recover the DNA. | |
| 8 | | Wash beads with freshly prepared 70% ethanol. Note that 70% ethanol is hygroscopic and should be prepared FRESH to achieve optimal results. Also, 70% ethanol should be stored in a tightly capped polypropylene tube for no more than 3 days. <ul style="list-style-type: none">– Do not remove the tube from the magnetic rack.– Use a sufficient volume of 70% ethanol to fill the tube (1.5 mL for 1.5 mL tube or 2 mL for 2 mL tube). Slowly dispense the 70% ethanol against the side of the tube opposite the beads.– Do not disturb the bead pellet.– After 30 seconds, pipette and discard the 70% ethanol. | |
| 9 | | Repeat step 8 above. | |
| 10 | | Remove residual 70% ethanol. <ul style="list-style-type: none">– Remove tube from magnetic bead rack and spin to pellet beads. Both the beads and any residual 70% ethanol will be at the bottom of the tube.– Place the tube back on magnetic bead rack.– Pipette off any remaining 70% ethanol. | |
| 11 | | Check for any remaining droplets in the tube. If droplets are present, repeat step 10 . | |
| 12 | | Remove the tube from the magnetic bead rack and allow beads to air-dry (with tube caps open) for 30 to 60 seconds. | |
| 13 | | For up to 10 µg input SMRTbell library, elute the DNA off the beads in 100 µL of Elution buffer. If you have more than 10 µg library, scale volume of EB proportionally. Add the Pacific Biosciences® Elution Buffer volume to your beads. Tap the tube with finger gently to mix. <u>Do not pipet to mix.</u> <ul style="list-style-type: none">– Elute the DNA by letting the mix stand at room temperature for 2 minutes.– Spin the tube down to pellet beads, then place the tube back on the magnetic bead rack.– Let beads separate fully. Then without disturbing the bead pellet, transfer supernatant to a new 1.5 ml Lo-Bind tube.– Discard the beads. | |
| 14 | | The eluted DNA in 100 µL Elution Buffer should be taken into the third 0.40X AMPure PB bead purification step. | |

| STEP | ✓ | Purify SMRTbell® Templates - Third Purification | Notes |
|------|---|---|-------|
| 1 | | Add 0.40X of AMPure PB beads to the eluted DNA. Note that for 0.40X volumes, it is critical to accurately pipet the desired volume of AMPure PB bead solution; there is a steep drop-off in recovery for concentrations < 0.40X . | |
| 2 | | Mix the bead/DNA solution thoroughly by gently tapping the tube. Do not pipet to mix. | |
| 3 | | Quickly spin down the tube (for 1 second) to collect the beads. Do not pellet beads. | |
| 4 | | Allow the DNA to bind to beads by gentle rotation for 15-20 minutes at room temperature. We recommend using a tube rotator. | |
| 5 | | Spin down the tube (for 1 second) to collect beads. | |
| 6 | | Place the tube in a magnetic bead rack to collect the beads to the side of the tube. | |
| 7 | | Slowly pipette off cleared supernatant and save (in another tube). Avoid disturbing the bead pellet. If the DNA is not recovered at the end of this procedure, you can add equal volumes of AMPure PB beads to the saved supernatant and repeat the AMPure PB bead purification steps to recover the DNA. | |
| 8 | | Wash beads with freshly prepared 70% ethanol. Note that 70% ethanol is hygroscopic and should be prepared FRESH to achieve optimal results. Also, 70% ethanol should be stored in a tightly capped polypropylene tube for no more than 3 days. <ul style="list-style-type: none"> – Do not remove the tube from the magnetic rack. – Use a sufficient volume of 70% ethanol to fill the tube (1.5 mL for 1.5 mL tube or 2 mL for 2 mL tube). Slowly dispense the 70% ethanol against the side of the tube opposite the beads. – Do not disturb the bead pellet. – After 30 seconds, pipette and discard the 70% ethanol. | |
| 9 | | Repeat step 8 above. | |
| 10 | | Remove residual 70% ethanol. <ul style="list-style-type: none"> – Remove tube from magnetic bead rack and spin to pellet beads. Both the beads and any residual 70% ethanol will be at the bottom of the tube. – Place the tube back on magnetic bead rack. – Pipette off any remaining 70% ethanol. | |
| 11 | | Check for any remaining droplets in the tube. If droplets are present, repeat step 10 . | |
| 12 | | Remove the tube from the magnetic bead rack and allow beads to air-dry (with tube caps open) for 30 to 60 seconds. | |
| 13 | | For up to 5 µg SMRTbell library, elute in 10 µL Elution buffer. For more than 5 µg of SMRTbell template, scale volume of EB proportionally (i.e., for up to 10 µg of input DNA, elute in 20 µL EB). Add the Pacific Biosciences® Elution Buffer volume to your beads. Tap the tube with finger gently to mix. Do not pipet to mix. <ul style="list-style-type: none"> – Elute the DNA by letting the mix stand at room temperature for 2 minutes. – Spin the tube down to pellet beads, then place the tube back on the magnetic bead rack. – Let beads separate fully. Then without disturbing the bead pellet, transfer supernatant to a new 1.5 ml Lo-Bind tube. – Discard the beads. | |

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|-----------|--|
| 14 | <p>Verify your DNA amount and concentration using a Qubit quantitation platform.</p> <ul style="list-style-type: none"> – Using 1 μL of the purified sample, make a 1:10 dilution in EB. – Use 1 μL of this 1:10 dilution to measure the DNA concentration using a Qubit fluorometer and the dsDNA HS Assay kit according to the manufacturer's recommendations. <p>The remaining 9 μL of 1:10 diluted sample may be used for QC by pulsed-field gel electrophoresis or pulse-field capillary electrophoresis.</p> <p>Note that typical DNA yield, at this point of the process (at the end of library preparation) is between approximately 5-20% of the total starting DNA amount.</p> |
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Primer Annealing, Polymerase Binding and Sequencing

Please refer to the Binding Calculator and SMRT Link Sample Setup for RSII and Sequel, respectively.

| Revision History (Description) | Version | Date |
|---|---------|--------------|
| Removed DNA Damage Repair requirement. Provided elution volume (page 3). Added step to check for droplets (in Purification tables). Removed loading recommendations. Changed the volume of blunt adapter from 1 μ L to 10 μ L. | 06 | January 2018 |
| ONLY CHANGE IS TO REVISION HISTORY BOX FROM VERSION 06: Concentrate DNA (page 3/step13): Provided Elution volume (37 μ L). Changed the volume of blunt adapter from 1 μ L to 10 μ L. On page 12, removed specifics related to annealing, binding, and sequencing and referred to Binding Calculator and SMRT Link Sample Setup. | 07 | January 2018 |

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