IsoPhase: Phasing Iso-Seq Data for diploid (and possibly tetraploid) genomes

Elizabeth Tseng, Jan 2018, PAG SMRT Developers Conference
Online Resources:

Google Groups: groups.google.com/forum/#!forum/SMRT_isoseq

GitHub: github.com/PacificBiosciences/IsoSeq_SA3nUP/
   (shortened: http://tinyurl.com/PBisoseq)

Twitter: @magdoll
<table>
<thead>
<tr>
<th>TYPE</th>
<th>SPECIES</th>
<th>TITLE</th>
</tr>
</thead>
</table>
PUBLICLY AVAILABLE SEQUEL ISO-SEQ DATA

**Sequel System Data Release: Iso-Seq Results for Hummingbird and Zebra Finch Brain Tissue**

Thursday, August 31, 2017

If you’re interested in avian vocal learning or want to explore a PacBio Iso-Seq data set generated with the Sequel System, we have good news. We’ve just released data from Iso-Seq interrogations of brain tissue from two avian models of vocal learning, Anna’s hummingbird (*Calypte anna*) and zebra finch (*Taeniopygia guttata*), sequenced in collaboration with the Erich Jarvis and Olivier Fedriga labs at the Rockefeller University.

If you’re not familiar with the Iso-Seq method, it’s the long-read sequencing answer to short-read RNA-seq studies. By using SMRT Sequencing for a transcriptome project, scientists can generate full-length isoform data, clearly capturing alternative splicing events to see the real diversity of transcripts. Unlike RNA-seq approaches, the Iso-Seq method takes advantage of long-read data to fully span transcript isoforms from the 5’ end to their poly-A tails, eliminating the need for error-prone transcript reconstruction and inference processes. With the Sequel System, Iso-Seq projects are low cost and time efficient. Currently we recommend only 1-2 SMRT Cells per tissue type for genome annotation.

- 4 Sequel cells
- Barcoded bird brains
- Total: 785k FL reads

<table>
<thead>
<tr>
<th></th>
<th>ZEBRAFINCH</th>
<th>HUMMINGBIRD</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Runtime</strong></td>
<td>31 hr</td>
<td>28 hr</td>
</tr>
<tr>
<td><strong>Unique Genes</strong></td>
<td>7228</td>
<td>7357</td>
</tr>
<tr>
<td><strong>Unique Isoforms</strong></td>
<td>17,437</td>
<td>16,898</td>
</tr>
</tbody>
</table>
ISO-SEQ2: COMING TO YOU IN 2018!

- Faster runtime
- Increased transcript recovery
- Reduced false artifacts
- Available in the next SMRTLink release

<table>
<thead>
<tr>
<th>SMRT CELLS</th>
<th>CCS READS</th>
<th>FL READS</th>
<th>ISOSEQ2 RUNTIME</th>
<th>UNIQUE TRANSCRIPTS</th>
<th>UNIQUE GENES</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>244,804</td>
<td>212,201</td>
<td>15 hr</td>
<td>13,036</td>
<td>7760</td>
</tr>
<tr>
<td>3</td>
<td>796,128</td>
<td>676,905</td>
<td>21 hr</td>
<td>30,956</td>
<td>13,044</td>
</tr>
<tr>
<td>6</td>
<td>1,908,507</td>
<td>1,562,039</td>
<td>5 days</td>
<td>63,645</td>
<td>18,227</td>
</tr>
</tbody>
</table>

* run using all default options except CCS minimum pass changed to 1
Phasing Iso-Seq Data
**MOTIVATION**

- The Iso-Seq bioinformatics pipeline outputs distinct isoforms (exon skipping, alternative 5’ and 3’ ends), but collapses SNP-level variations.

- SNP information can be revealed by aligning full-length (FL) CCS reads back to the unique isoforms after Iso-Seq analysis.
"quickphase" in IGV separates aligned reads into two groups
MOTIVATION

- Juliet calls minor variants in viral data
- However,
  - It does not handle splicing
  - Performs best with stringent CCS cutoff (> 99%)
  - Performs best with deep coverage (> 250-fold)
ISOPHASE: ISOFORM PHASING USING ISO-SEQ DATA

ALIGNMENT

SNP CALLING

Position | SNPs
---------|------
POS1     | A, G
POS2     | C, T
POS3     | C, A

PHASING

VCF OUTPUT

```plaintext
##fileformat=VCFv4.2
#CHROM  POS ID  REF ALT  QUAL  FILTER  INFO    FORMAT  ISOFORM1  ISOFORM2
chr1   105 .   A   G   .   PASS    DP=40;AF=0.50   GT:HQ   0|1:20,20 0:15
chr1   190 .   C   T   .   PASS    DP=40;AF=0.50   GT:HQ   0|1:20,20 0:15
chr1   336 .   C   A   .   PASS    DP=40;AF=0.50   GT:HQ   0|1:20,20 0:15
```
ISOPHASE METHOD SUMMARY

- Alignment using minimap2, retrieve positions with sufficient coverage
  - If QV is provided, alignment pileup filters out low-quality bases

- SNP calling and phasing using Juliet

- Simple clustering of phased haplotypes to remove errors

- Output VCF denoting SNPs and allele counts for each isoform
SNP CALLING AND PHASING IN JULIET

- Steps across alignment in codon triples. *not applicable for transcriptome*
- Computes a p-value that the observed bases come from purely noise using a Fisher’s exact test.
- If the p-value is significant under a Bonferroni correction, call the variant.
- Phase together variant positions by tallying full-length reads that exhibit different combinations of the variant positions and threshold.
- Limits:
  - CCS reads used to minimize impact of noise. Raw read analysis possible.
  - Currently does not estimate indel variants.
- Reliably identify 1% true variants from sequencing noise.
Evaluation on Simulated Data
SNP CALLING ON SIMULATED 100 HUMAN GENES

- Random 100 human genes
- Simulated 1 SNP per 300 bp
- Each allele has 20 copies (20-fold coverage)
- Simulated substitution errors at 1%, 2%, 5%, and 10%

![Line graph showing True Positive Rate vs Simulated Error Rate for different types of sequencing reads. The graph indicates that as the simulated error rate increases, the true positive rate decreases for both types of reads, with 2N-withQV showing slightly better performance than 2N-noQV.](image)
SNP CALLING ON SIMULATED 100 HUMAN GENES

- Using QV improves SNP recovery
- SNP recovery (TPR) remains high for both diploid and tetraploid even at 10% error
SNP CALLING ON SIMULATED 100 HUMAN GENES

- False discovery rate remains low until 5% error
- False discovery rate increases with error rate for tetraploid-noQV
PHASING EVALUATION: CRITERION

For 4N, “correct phasing” means getting all 4 alleles correct. Getting 3 → still wrong.
PHASING EVALUATION ON SIMULATED 100 HUMAN GENES

Percentage of 100 genes that were correctly phased.

<table>
<thead>
<tr>
<th>Error Rate</th>
<th>2N no QV</th>
<th>2N with QV</th>
<th>4N no QV</th>
<th>4N with QV</th>
</tr>
</thead>
<tbody>
<tr>
<td>1%</td>
<td>100%</td>
<td>100%</td>
<td>95%</td>
<td>98%</td>
</tr>
<tr>
<td>2%</td>
<td>100%</td>
<td>100%</td>
<td>82%</td>
<td>98%</td>
</tr>
<tr>
<td>5%</td>
<td>100%</td>
<td>100%</td>
<td>55%</td>
<td>96%</td>
</tr>
<tr>
<td>10%</td>
<td>100%</td>
<td>93%</td>
<td>40%</td>
<td>84%</td>
</tr>
</tbody>
</table>

Worse than no QV due to aggressive dropping of reads
(future work: relax criterion for using reads in haplotyping)
POTENTIAL TO RECOVER ALLELIC SPECIFIC EXPRESSION

Ratio of most abundant : least abundant allele.

Except for (20:40), all 2N simulated with (20:20) read coverage.
All 4N simulated with (20:20:20:20) read coverage.
Evaluation on F1 Cattle Data
ANGUS X BRAHMAN F1 CATTLE

Genome Assembly
- Angus (sire) x Brahman (dam) F1 cattle
- 115-fold coverage on PacBio RS II and Sequel systems
- Assembled using Falcon
- ~90% of genome phased using Unzip

<table>
<thead>
<tr>
<th>CONTIG</th>
<th>NUMBER</th>
<th>LENGTH</th>
<th>N50</th>
<th>LONGEST</th>
</tr>
</thead>
<tbody>
<tr>
<td>PRIMARY</td>
<td>1427</td>
<td>2.71 Gb</td>
<td>31.4 Mb</td>
<td>65.3 Mb</td>
</tr>
<tr>
<td>HAPLOTIGS</td>
<td>5879</td>
<td>2.45 Gb</td>
<td>2.48 Mb</td>
<td>14.0 Mb</td>
</tr>
</tbody>
</table>

Iso-Seq Transcriptome Data
- 8 Sequel cells of tissues from single individual
- Analyzed using IsoSeq2
- Mapped to genome with ≥ 99% coverage, ≥ 95% identity
- 30,137 final isoforms (12,101 genes)
- Selected for phasing: 1758 genes with ≥ 40 full-length CCS read coverage
SNP EVALUATION FOR ANGUS X BRAHMAN

<table>
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<tr>
<th>SNP Type</th>
<th>Count</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>True Positive</strong> (called by both)</td>
<td>8334</td>
</tr>
<tr>
<td><strong>False Negative</strong> (called by genome only)</td>
<td>259</td>
</tr>
<tr>
<td><strong>Unphased by Genome</strong> (called by transcript only)</td>
<td>1203</td>
</tr>
</tbody>
</table>

Using genome phasing results as truth, IsoPhase SNP calling achieves 97% sensitivity and 87% specificity.
EXAMPLE OF SNP CALLING VERIFIED BY GENOME

Full-length CCS read alignment, showing only exons with SNPs. Reads are sorted through “quickphase” in IGV browser showing clear segregation of alleles. All 6 SNPs validated by genome assembly Unzip results.
EXAMPLE OF SNP CALLING VERIFIED BY GENOME

There are 5 different isoforms for this gene. All isoforms cover all 6 SNP sites.
This gene (PB.1001, VPS36) contains 228 FL reads.

- Strong evidence for the 3 SNPs.
- Unzip did not phase this region – so, are the SNPs supported by genome?
VPS36 ISOFORMS CALLED SNPS NOT PHASED IN GENOME

The first SNP 000004F|arrow|arrow:48163477 (C->G) is supported in the pre-polish BAM file.
VPS36 ISOFORMS CALLED SNPS NOT PHASED IN GENOME

The second SNP 000004F|arrow|arrow:48163716 (T→G) is the second T in the sequence context GCCCGGACCT\_TGCCCATAGG which in the pre-polish BAM file shows evidence that the second “T” is either a “T” or a “G”.

the insertion is either a “T” or a “G” which is the SNP
VPS36 ISOFORMS CALLED SNPS NOT PHASED IN GENOME

The third SNP 000004F|arrow|arrow:48166508 (A->T) is also an insertion against the pre-polish sequence that is supported by the genome subread data.

the insertion is either a “A” or a “T” which is the SNP
POTENTIAL A → G RNA EDITING IN COL1A1

<table>
<thead>
<tr>
<th>CHROM</th>
<th>POS</th>
<th>REF</th>
<th>ALT</th>
<th>SNP IN GENOME?</th>
</tr>
</thead>
<tbody>
<tr>
<td>000071F</td>
<td>7663000</td>
<td>A</td>
<td>G</td>
<td>N</td>
</tr>
<tr>
<td>000071F</td>
<td>7671641</td>
<td>T</td>
<td>C</td>
<td>Y</td>
</tr>
</tbody>
</table>

PB.8679 gene (COL1A1) contains a A → G SNP not supported by genome. A single alternative contig (000071F_029) covers the whole region.
POTENTIAL A → G RNA EDITING IN COL1A1

A → G SNP site

Alignment (3429 reads)

Isoforms
Conclusion: COL1A1 contains an non-dominant isoform (PB.8679.1) that uses an alternative donor splice site in exon 5 that includes a potential A → G editing site.
• KIF3C is observed in brain only
• The SNP is in the 3’ UTR region (A → G) and is verified by genome
• The major isoform expresses the A allele more dominantly
## SNP EVALUATION FOR BRAHMAN X ANGUS

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Using genome phasing results as truth, IsoPhase SNP calling achieves 97% sensitivity and 87% specificity.

However, many of the transcript-only SNPs could be true. They could be not phased by the genome due to low heterozygosity, low coverage, or RNA editing.
ISOPHASE SUMMARY

IsoPhase is a direct extension of the Iso-Seq analysis, utilizing full-length read information to detect SNPs and call haplotypes.

Based on both simulated and real data, it shows high true discovery rate and low false positive rate.

It has the potential to reveal allelic specific isoform expressions.
FUTURE WORK

Detect Indels
- Currently, only substitution SNPs are called
- Calling simple (1-3 bp) indels is conceptually possible, but will require work

Reduce Read Coverage Requirement + Short Read Support
- Currently, requires 40-fold per-gene read coverage
- Reducing read coverage may increase detection but also false calls
- Include short read data for SNP calling; use long reads for phasing

Phasing Without a Reference Genome
- Cogent could be used to reconstruct coding “contigs” to map full-length reads back to. However Cogent will require minor modifications to understand unresolved exonic orderings based solely on Iso-Seq data.

Pipeline for Comparing Genome Phasing Results with IsoPhase
- Automated scripts for showing agreement and disagreement

Please let me know if you have other ideas for making IsoPhase more awesome!