



Targeted long-read sequencing of native DNA for genetic disease diagnostic and screening research

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Introduction

Short tandem repeats (STRs) are DNA sequences composed of repetitions of 2 – 6 bp motifs. Expansions of STRs are the cause of over 60 monogenic diseases, including Huntington's disease, fragile X syndrome, and amyotrophic lateral sclerosis¹. In addition to their length, the pathogenicity of these STRs can be impacted by sequence composition, methylation status and mosaicism. One such example is the FMR1 repeat whose CGG repeat expansions are typically hypermethylated and where AGG interruption sequences can stabilize the repeat. Detecting all the characteristics associated with pathogenic repeat expansions traditionally required multiple assays, however high-accuracy long-read sequencing of unamplified DNA can resolve all these features in a single assay.

Scalable amplification-free workflow

- High molecular weight DNA
- 50% fragments > 30 kb
- 1 – 4 µg per sample

- Dephosphorylate to block DNA ends

- Cas9 cut with pair of sgRNAs
- "normal" length = 4 – 5 kb

- dA tail cut ends

- Ligate indexed SMRTbell adapters

- Nuclease digestion of non-SMRTbell templates

- Pool and sequence up to 48 samples on Revio or Vega system

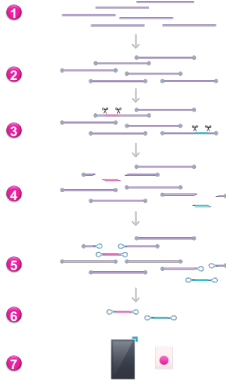


Figure 1. PureTarget repeat expansion panel is a robust amplification-free approach to generate long-read HIFI sequencing libraries containing loci associated with 20 pathogenic STR expansions. Starting with high molecular weight DNA from blood or cell line extracted with Nanobind PanDNA kit, the workflow employs Cas9 and a single pair of guide RNAs to target each repeat region and ~3-5 kb of flanking sequence. Comprehensive genotyping of consensus repeat size, motif analysis and methylation is performed with Tandem Repeat Genotyping Tool (TRGT)² in SMRT Link software.

Gene(s)	Associated disease
ATM1, ATXN1, ATXN2, ATXN3, ATXN7, ATXN8, ATXN10, CACNA1A, PPP2R2B, TBP	Spinocerebellar ataxia
FMR1	Fragile X-associated disorders
C9orf72	Amyotrophic lateral sclerosis and Frontotemporal dementia
DMPK, CNBP	Mitochondrial dystrophy (DM1, DM2)
FXN	Friedreich's ataxia
RFC1	CANVAS
HTT	Huntington's disease
AR	Spinal-bulbar muscular atrophy
PABPN1	Oculopharyngeal muscular dystrophy
TCF4	Fuchs endothelial corneal dystrophy

Table 1. PureTarget Repeat Expansion Panel Content. The PureTarget product line includes a panel of 20 repeat expansion loci relevant to neurodegenerative disease.

References and Resources

- Leitão, E., et al. (2024). Identification and characterization of repeat expansions in neurological disorders: Methodologies, tools, and strategies. *Rev Neurol (Paris)*, 180(5):383-392. doi: 10.1016/j.neuro.2024.03.005.
- Dobzhenko, E., et al. (2024). Characterization and visualization of tandem repeats at genome scale. *Nat Biotechnol*, 2024. doi: 10.1038/s41587-023-02057-3.



Datasets description

To assess the accuracy of PureTarget across sequencing platforms and chemistries, we sequenced panels of reference samples with validated pathogenic expansions. By including the same sample across multiple panels, and sequencing each pool across multiple platforms, we allow for both technical replicates and robust comparison across the sequencing technologies.

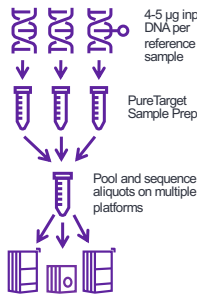


Figure 2 (left). Diagram of the experimental set up. We prepared 4-5 µg of input DNA with PureTarget for each sample. Libraries were pooled and sequenced on both technologies to allow for comparison of both technical and sequencing replicates.

Table 2 (below). List of panels generated for these results. Sample pools were composed of 16 or 24 samples, with potential technical replicates within a panel. As shown in Figure 2, pooled libraries were aliquoted across sequencers. We aliquoted such that Revio v1 and Vega received 2µg input DNA per sample equivalent, while Revio system with SPRQ chemistry received 1 µg of input DNA, thanks to ability to load lower input samples.

Panel	Plex	Vega	Revio v1	Revio SPRQ
Coriell A	24*	2µg	2µg	1µg
Coriell B	24	2µg	2µg	N/A
Coriell C	16	2µg	2µg	1µg

*Two samples from Coriell A are excluded from this report due to insufficient input material (< 2µg instead of 5µg).

Sample coverage across platforms

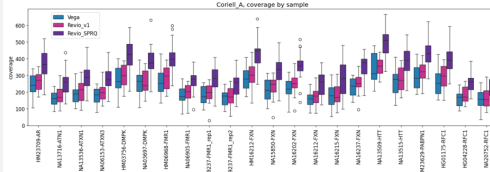


Figure 3. Target coverage by sample for reference Coriell samples with known repeat expansions (N=22) across different sequencing technologies. Target coverage is similar between Vega and Revio v1, while Revio SPRQ has higher coverage. Full dataset available at <https://downloads.pacbcloud.com/public/2024Q4/Vega/PureTargetCoriell24/>

Complex motif and mosaicism at HTT

A. Genotypes
Expected genotype: Short allele – (CAG)¹⁵
Long allele – (CAG)⁷⁸
Observed genotype: Short allele – (CAG)¹⁵(CAA)(CAG)(CCG)(CCA)(CCG)¹⁸
Long allele – (CAG)⁷³(CAA)(CAG)(CCG)(CCA)(CCG)⁷

B. Coverage and mosaicism
Vega Coverage
Short allele – 74-fold
Long allele – 95-fold
Revio Coverage
Short allele – 248-fold
Long allele – 247-fold

Vega Mosaicism
Short allele – 83-91
Long allele – 237-274

Revio Mosaicism
Short allele – 84-97
Long allele – 234-282

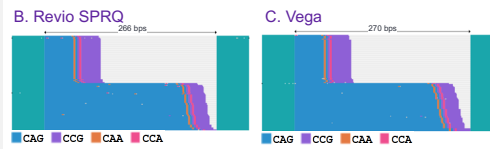


Figure 4. Six technical replicates of sample NA13509, from an individual clinically affected by Huntington's disease, were sequenced giving consistent genotypes (A) across platforms. Waterfall plots down sampled to 50 reads per allele for Revio SPRQ (C) and Vega (D) reveal high depth and low level of mosaicism in the long allele, ranging from ~234 – 282 bps on Revio (B).

Mosaicism at FXN

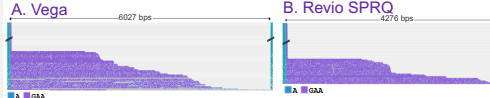


Figure 5. Waterfall plots for NA16237 at the FXN locus, sequenced on Vega (A) and Revio SPRQ (B). The short allele does not show mosaicism and was partially cropped from the waterfall plots. The long allele shows high mosaicism with alleles ranging from ~2000 to ~5000 bps. Coverage (long/short) for Revio SPRQ was 200 / 29 reads and for Vega was 131 / 37.

Expanded alleles in technical replicates

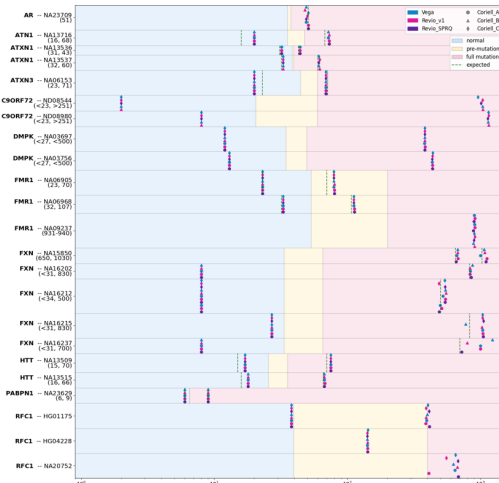


Figure 6. Swimlane plot showing the genotype calls for replicates of the reference samples. Samples with 3 or more replicates are shown. Color indicates the sequencing platform, while shape represents the technical replicate. Points with the same shape come from the same original aliquot. Allele length is reported as the number of pathogenic repeat units. When available, the expected genotype is specified on the y-axis label. Background colors indicate whether the length is considered normal, pre-mutation or full mutation¹.

Genotyping at C9orf72 on Vega

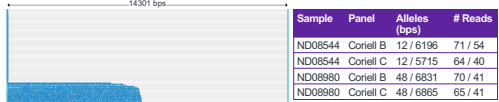


Figure 7. Genotyping calls and read coverage for C9orf72 expansions on Vega. Technical replicates were sequenced on Vega for two samples with known C9orf72 expansions, ND08544 and ND08980. For both samples, we see consistent coverage of the expanded allele, which contains repeats > 5000 bps for ND08544 and > 6500 bps for ND08980. The long allele shows mosaicism. Read coverage was > 40 for both allele on all samples and replicates.

Methylation and AGG motif at FMR1

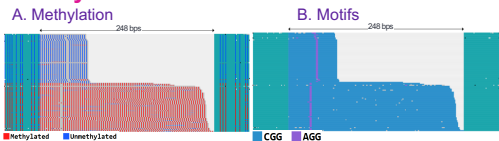


Figure 8. FMR1 waterfall plots on Vega. Coverage is downsampled to 100 reads. When representing methylation (A), we note that the short allele is unmethylated, while the long allele is hypermethylated. Looking at the motif sequence, we can see that both alleles contain the protective AGG interruptive motif.

Conclusions

- PureTarget is a **complete solution** to accurately characterize lengths, repeat sequence and methylation status of repeat expansions relevant for human disease.
- The PureTarget repeat expansion panel protocol and analysis in SMRT Link can deliver **sample to answer in 3 days**.
- PureTarget delivers **consistent results across technical replicates and HIFI sequencing platforms** for reference samples with known repeat expansions.